

Analysis protocol for GENE-SWitCH capture Hi-C data (pig and chicken)

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Pig and chicken GENE-SWitCH capture Hi-C data (*in situ* Hi-C followed by capture enrichment and high throughput sequencing) were processed using the nf-core hic pipeline (<https://github.com/nf-core/hic>) version 1.3.0.

For pig and using a slurm cluster, we performed the following steps:

1) Input data

From hereafter the species name, either <sus_scrofa> for pig or <gallus_gallus> for chicken, will be referred to as `$species`.

Input data are stored in a <data.\$species> directory

- a) Get raw sequencing reads in fastq.gz format from the ENA
 - Pig: Accessions PRJEB44198 and PRJEB44486
 - Chicken: Accessions PRJEB53986 and PRJEB54773
- b) Get the genomic sequences ready (here downloaded from ENSEMBL build 102)
 - Pig: sus_scrofa.fa (Sscrofa11.1)
 - Chicken: gallus_gallus (GRCg6a)
- c) Get the corresponding bowtie2 indices, named after the genome file
\$species.fa.*.bt2 (Bowtie2 version: 2.3.5.1)
with the command

```
bowtie --index $species.fa
bowtie2-build $species.fa $species.fa
```
- d) Get a file with the chromosomes and contigs' lengths using for instance the fastalength tool from the Exonerate package version 2.2.0

```
fastalength $species.fa | awk '{print $2"\t"$1}' >
chrom.len
```

2) Launch the nf-core hic on the GENE-SWitCH capture Hi-C reads

- a) make a config file `capturehic.conf`

```
singularity {
  enabled = true
}
process {
  withName:bowtie2_end_to_end {
    cpus = 24
    memory = 64.GB
```

```

    time = 36.h
  }
  withName:trim_reads {
    memory = 32.GB
    time = 72.h
  }
  withName:bowtie2_on_trimmed_reads {
    cpus = 16
    memory = 64.GB
    time = 36.h
  }
  withName:bowtie2_merge_mapping_steps {
    cpus = 8
    time = 36.h
  }
  withName:get_valid_interaction {
    memory = 64.GB
    time = 36.h
  }
  withName:remove_duplicates {
    cpus = 32
    memory = 160.GB
    time = 36.h
  }
  withName:tads_hicexplorer {
    memory = 64.GB
    time = 36.h
  }
  withName:tads_insulation {
    memory = 32.GB
    time = 36.h
  }
  withName:dist_decay {
    memory = 64.GB
    time = 36.h
  }
  withName:compartment_calling {
    memory = 32.GB
    time = 36.h
  }
  withName:combine_mates {
    memory = 16.GB
  }
  withName:merge_stats {

```

```

        time = 8.h
    }
    withName:multiqc {
        memory = 8.GB
        time = 3.h
    }
    withName:build_contact_maps {
        memory = 32.GB
        time = 6.h
    }
}

```

- b) make a bash script called `capturehic.$species.sh` to be launched (here on a slurm cluster) with the following content:

```

#!/bin/bash
mkdir -p capturehic.$species
cd capturehic.$species

module load bioinfo/nfcore-Nextflow-v21.10.6
nextflow run nf-core/hic \
  -revision 1.3.0 \
  -profile genotoul \
  -resume \
  -c capturehic.conf \
  -work-dir capturehic.$species/work \
  --fasta data.$species/$species.fa \
  --bwt2_index data.$species \
  --outdir capturehic.$species \
  --input 'data.$species/*_R{1,2}.fastq.gz' \
  --bwt2_opts_end2end '-5 5 -3 95 --very-sensitive -L 20 --score-min L,-0.6,-0.3 --end-to-end --reorder' \
  --bwt2_opts_trimmed '--very-sensitive -L 20 --score-min L,-0.6,-0.3 --end-to-end --reorder' \
  --min_mapq 10 \
  --restriction_site '^GATC,G^ANTC' \
  --ligation_site 'GATCGATC,GANTGATC,GANTANTC,GATCANTC' \
  --chromosome_size data.$species/chrom.len \
  --min_insert_size 10 \
  --max_insert_size 1000 \
  --bin_size '500000,10000,5000,1000' \
  --ice_max_iter 100 \
  --ice_filter_low_count_perc 0.02 \
  --ice_filter_high_count_perc 0 \

```

```
--ice_eps      0.1 \
--res_compartments '500000,200000' \
--tads_caller    'insulation,hicexplorer' \
--res_tads      '100000,50000' \
--save_interaction_bam \
--skip_balancing \
--skip_tads
```

c) *launch it on a slurm cluster with the command*

```
sbatch capturehic.$species.sh
```

3) Submitted analysis files

a) *Hi-C interaction matrices (contact maps)*

Those are contact matrices in .cool format, one per fastq.gz read pair, that were located in the capturehic.\$species/contact_maps/raw/cool directory