Standard operating procedure for single cell isolation from fresh bovine liver tissue

modified after the protocol from Ramachandran P. et al. (2019) (PMID: 31597160)

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Purpose

Collecting single cells from fresh bovine liver tissue. Appropriate preparation of required materials and professional training of lab technicians involved cell isolation have to be performed in advance.

Materials

- pipettes, pipette tips
- 1X PBS
- 1X PBS + 0.04% BSA
- beakers
- 1X HBSS
- enzyme cocktail (5 mg/ml Pronase, 2.93 mg/ml Collagenase B and 1.9 mg/ml DNase I)
- heater shaker
- 50 ml Falcon tubes
- 100 μm cell strainer
- PEB buffer (1X PBS, 1 mg/ml BSA, 2 mM EDTA)
- DNase I
- Red Blood Cell lysis buffer
- centrifuge, table centrifuge
- AO/PI
- Cellometer Auto2000

Single-cell isolation

- 1. the animals are killed according to regular slaughtering protocol including stunning and exsanguination by expert staff in an experimental slaughterhouse of the institute
- 2. liver tissue is collected during slaughter by expert staff and distributed to trained lab technicians for further sample preparation
- 3. liver tissue is washed with cold 1X PBS
- 4. a walnut-sized piece of liver is cut from the tissue (without capsule) and placed in a beaker with cold 1X HBSS (for transport to the laboratory)
- 5. tissue is minced into small pieces (1-2mm2).
- 6. tissue pieces are incubated in a 50 ml Falcon in 10 ml enzyme cocktail of 5 mg/ml Pronase, 2.93 mg/ml Collagenase B and 1.9 mg/ml DNase I at 37°C for 30 min with gentle shaking (200 rpm)
- 7. remaining tissue pieces are removed
- 8. a 100 μ m cell filter is placed on a 50 ml Falcon and the cell suspension is filtered using PEB buffer (1X PBS, 1 mg/ml BSA, 2 mM EDTA) and 0.02 mg/ml DNase I (if necessary: use a plunger of a disposable syringe to support the filtering of the cell suspension by rubbing on the filter)

- 9. Falcon is centrifuged with 400 g for 7 min at 4°C and the supernatant is removed
- 10. cells are resuspended in 20 ml cold (1X) Red Blood Cells lysis solution and the Falcon is inverted several times
- 11. cells are incubated in 1X RBC lysis solution for 5 min at room temperature
- 12. Falson is centrifuged with 400 g for 7 min at 4°C and the supernatant is removed
- 13. cells are resuspended in 15 ml PEB buffer with 0.02 mg/ml DNase
- 14. cells are centrifuged with 400 g for 7 min at 4°C and the supernatant is removed
- 15. cells are resuspended in 5 ml 1X PBS + 0.04% BSA
- 16. cell concentration and viability are measured using AO/PI staining and a Cellometer Auto2000 (Nexcelom)