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1. Description

This product is for research use only.

Components 100 mL Nuclei Extraction Buffer

Size For 25 extractions.

Storage Store protected from light at 2–8 °C. Do not freeze. The expiration date is indicated on the label.

1.1 Principle of the Nuclei Extraction Buffer

Tissue samples from various sources and species can be dissociated into single nuclei suspensions by combining mechanical tissue dissociation and cell lysis. The tissue sample is added to a gentleMACS™ C Tube together with the Nuclei Extraction Buffer and RNase inhibitor. The sample is dissociated using the gentleMACS Dissociator.

After dissociation, the sample is applied to a filter to remove any remaining larger particles from the single-nuclei suspension. Nuclei are collected by centrifugation and the nuclei pellet is resuspended and filtered again.

1.2 Background information

The Nuclei Extraction Buffer and the corresponding gentleMACS Program have been optimized for a gentle, rapid, and effective generation of single-nuclei suspensions from fresh and frozen tissue samples. This reliable and standardized protocol enables the recovery of high yield single-nuclei suspensions that can be used for a wide variety of studies including proteomic analysis, FACS analysis, and single-nuclei gene expression analysis.

1.3 Applications

- Dissociation of fresh, frozen (including flash-frozen and OCT embedded) tissue samples from various tissues and species into single-nuclei suspensions for subsequent molecular analyses.
- The provided protocol has been validated for a wide range of tissues, including brain, liver, heart, lung, pancreas, spleen and kidney, as well as tumor tissues including human melanoma,

breast, pancreatic, colon and prostate cancer and grafted mouse tumors.

1.4 Reagent and instrument requirements

- Centrifuge
- MACS® SmartStrainers (70 µm) (# 130-098-462)
- MACS SmartStrainers (30 µm) (# 130-098-458)
- gentleMACS™ Octo Dissociator (# 130-095-937), gentleMACS Octo Dissociator with Heaters (# 130-096-427)
- gentleMACS C Tubes (# 130-093-237, # 130-096-334)
- RNase-Inhibitor (e.g. murine RNase Inhibitor from New England Biolabs)
- (Optional) Eppendorf® LoBind® Tubes 5 mL

Additional requirements for working with mouse brain

- MACS SmartStrainers (100 µm) (# 130-098-463)

Additional requirements for microscopic or flow cytometric analysis of single-nuclei suspensions (refer to protocol 2.3)

- Nuclei staining dye, e.g. 7-AAD Staining Solution (# 130-111-568), DAPI Staining Solution (# 130-111-570), or DRAQ5 Staining Solution (# 130-117-343).
- (Optional) Cell sorter, e.g. MACSQuant® Tyto® Sorter (# 130-103-931)

2. Protocol

▲ For details on the use of the gentleMACS Dissociators, refer to the gentleMACS Dissociators user manuals.

▲ Please make sure that your gentleMACS Octo Dissociator runs with the latest software update. The **4C_nuclei_1** program is included in the Miltenyi program folder from software version GM_V02.H21 (gentleMACS Octo Dissociator with Heaters) or GM_V02.R11 (gentleMACS Octo Dissociator) onwards. The latest software update can be requested under <https://www.miltenyibiotec.com/ly/2018/gentlemacs-software-update.html>.

▲ Pipette gently and slowly during all nuclei resuspension steps to minimize alteration of the extracted nuclei.

2.1 Sample, equipment, and reagent preparation

▲ Pre-cool centrifuge, buffers, and consumables with sample contact (e.g. gentleMACS C Tube and SmartStrainers) at 4 °C.

▲ **Note:** It is recommended to pre-cool the C Tube containing prepared lysis buffer overnight at 4 °C. Ideally, perform all nuclei extraction steps in a cold room.

Preparation of lysis buffer

Per extraction add RNase inhibitor (final concentration 0.2 U/μL) to pre-cooled 4 mL Nuclei Extraction Buffer.

Preparation of resuspension buffer

Prepare an appropriate amount of resuspension buffer, e.g. PBS with 0.1% BSA and 0.2 U/μL RNase inhibitor. Store at 4 °C. Per extraction the following volumes of resuspension buffer are recommended:

Amount/type of tissue	Resuspension buffer
up to 50 mg	1.5 mL
50–100 mg	3 mL
100–200 mg	6 mL
Mouse brain hemisphere	8 mL

Table 1: Overview of resuspension volumes.

▲ **Note:** It is strongly recommended to adjust the nuclei concentration to < 5×10⁶/mL to avoid nuclei aggregation.

▲ **Note:** When working with tissues with high intrinsic RNase activity (e.g. pancreas), the RNase inhibitor concentration can be increased in both prepared lysis and resuspension buffer, in order to prevent RNA degradation.

▲ **Note:** If only DNA-based downstream applications are to be performed, RNase inhibitor can be left out.

▲ **Note:** Some genomic applications require the use of different resuspension buffer to ensure optimal performance. Please, follow the recommendations of the genomic application provider.

Preparation of tissue sample

Resect tissue and cut into pieces of up to 200 mg. To store tissue, freeze them on dry ice or use liquid nitrogen.

▲ **Note:** It is recommended to cut tissue before freezing. If frozen tissue pieces are > 200 mg, cut them without letting samples thaw.

▲ **Note:** In minor cases, e.g. kidney, it is recommended to cut up the tissue into pieces of < 50 mg to ensure complete dissociation.

▲ **Note:** When working with OCT-embedded tissue, remove excess OCT without letting samples thaw.

2.2 Nuclei extraction protocol

▲ Please consider that optimal extraction performance depends on starting material (tissue type, amount of starting material) and downstream application and might require protocol optimization.

▲ Work fast until tissue is dissociated and keep samples on ice at all steps.

▲ Volumes given below are for up to 200 mg tissue per extraction.

1. Add 2 mL ice-cold lysis buffer to each pre-cooled gentleMACS C Tube.
2. Transfer tissue pieces to the gentleMACS C Tube containing lysis buffer and directly proceed the following steps until samples are dissociated.

▲ **Note:** Do not let frozen sample thaw before dissociation as endogenous RNase might degrade RNA.

3. Close gentleMACS C Tube and place it on the gentleMACS Dissociator.

▲ **Note:** Close C Tube tightly beyond the first resistance.

4. Run gentleMACS Program **4C_nuclei_1** on the gentleMACS Dissociator.

5. After termination of the program, detach C Tube from the gentleMACS Dissociator and place the C Tube immediately on ice.

▲ **Note:** Depending on tissue type, additional 5-10 minutes incubation on ice after dissociation might be needed to optimize cell lysis. Over-lysis should be avoided as it could lead to nuclei aggregation and nuclei damage.

6. Apply nuclei suspension to a MACS SmartStrainer (70 μm) placed on a 15 mL tube.

▲ **Note:** For brain samples, use a MACS SmartStrainer (100 μm).

▲ **Note:** To increase nuclei recovery, the use of low binding tubes is recommended at this and all following steps.

7. Wash MACS SmartStrainer with 2 mL ice-cold lysis buffer.

▲ **Note:** Alternatively, for maximum cell recovery, rinse used C Tube with ice-cold lysis buffer before adding to the MACS SmartStrainer.

8. Discard MACS SmartStrainer and centrifuge nuclei suspension at 300×g at 4 °C for 5 minutes. Carefully aspirate supernatant completely.

9. Resuspend nuclei pellet with ice-cold resuspension buffer by slowly and gently pipetting the sample up and down for 10 times.

10. Apply nuclei suspension to a MACS SmartStrainer (30 μm) placed on a 15 mL tube.

▲ **Note:** When working with myelin-rich brain, a myelin removal step is strongly recommended to remove myelin debris.

11. Collect nuclei suspension and proceed immediately with downstream application, e.g., single nuclei RNA seq or nuclei sorting.

▲ **Note:** In case debris amount is still too high for downstream application, purify the nuclei suspension by using a cell sorter, e.g. MACSQuant Tyto.

2.3 Labeling for microscopic or flow cytometric analysis of nuclei samples

1. Add 7-AAD (final concentration of 0.525 μg/mL) or DAPI (final concentration of 0.25 μg/mL) Staining Solution or DRAQ5 (10 μM) Staining Solution to a small fraction of the nuclei suspension. Mix by inverting gently.
2. Incubate 5 minutes at 4 °C.
3. Load sample on a hemocytometer or on a flow cytometer, e.g. MACSQuant Analyzer.
4. Analyze sample according to manufacturer's recommendation.

Refer to www.miltenyibiotec.com for all data sheets and protocols. Miltenyi Biotec provides technical support worldwide. Visit www.miltenyibiotec.com for local Miltenyi Biotec Technical Support contact information.

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